PRECISION BIOSCIENCES

ARCUS nuclease-mediated excision of the "Hot Spot" region of the human dystrophin gene for the treatment of Duchenne Muscular Dystrophy (DMD)

American Society of Gene and Cell Therapy Session: Late-Breaking Abstracts 2 Gary Owens May 19, 2023



Forward-Looking Statements

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All forward-looking statements speak only as of the date of this presentation and, except as required by applicable law, we have no obligation to update or revise any forward-looking statements contained herein, whether as a result of any new information, future events, changed circumstances or otherwise.



Disclosures

• I am an employee of Precision BioSciences, Inc. (Nasdaq: DTIL)



Duchenne Muscular Dystrophy Currently Lacks a Curative Treatment

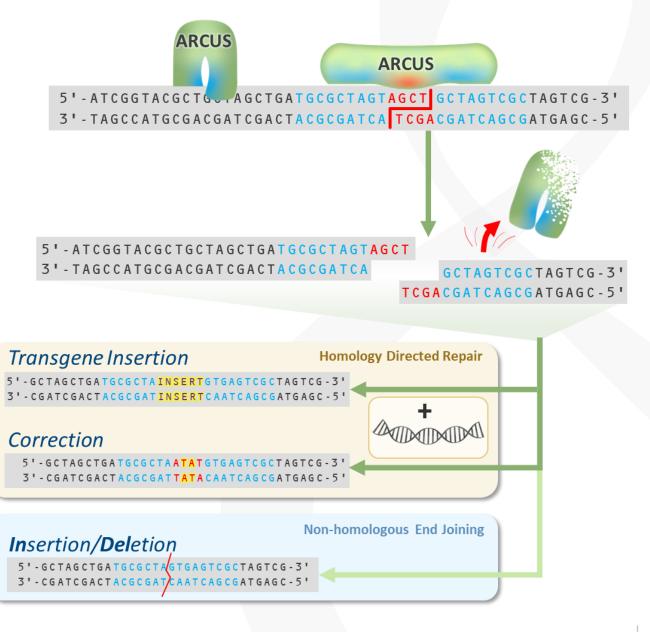




ARCUS: Engineering Nature's Genome Editing System

ARCUS

- Derived from I-CreI, a naturally-occurring green algae homing endonuclease
- Target site recognition and cleavage rely solely on an extensive DNA-protein interface
- DNA cleavage results in 4 bp 3' sticky ends
- Small size (364 aa) facilitates delivery via AAV or LNP





Utilizing ARCUS Nucleases to Restore Dystrophin Expression for DMD

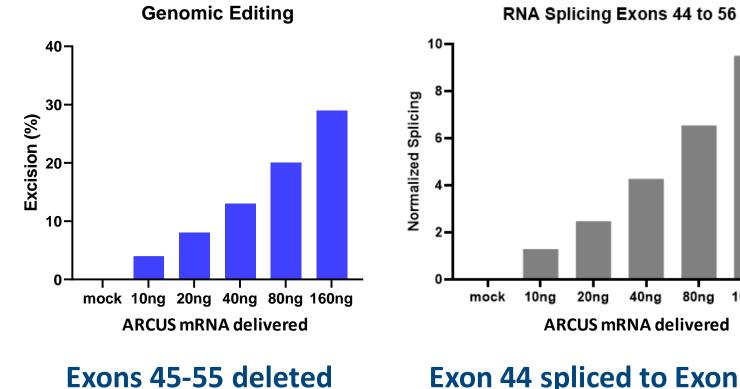
Strategy to restore dystrophin expression

Delete exons 45-55 using a pair of ARCUS nucleases intended to remove a mutation hotspot responsible for approximately 50% of DMD cases

ARCUS 1 ARCUS 2 GOAL Exons 45-55 Exon 44 Exon 56 Exon 57 Two complementary ARCUS nucleases Exons 45–55 deletion delivered in a single AAV are used to Exon 56 Exon 57 Exon 44 make a complex edit of the genome which results in the generation of a genome repair via direct re-ligation "Becker's like," functionally competent Exon 44 Exon 56 Exon 57 variant of the dystrophin protein intron splicing mRNA Exon 56 Exon 44 Exon 57 🔰



Dystrophin Gene Correction Observed in DMD Patient Myoblasts



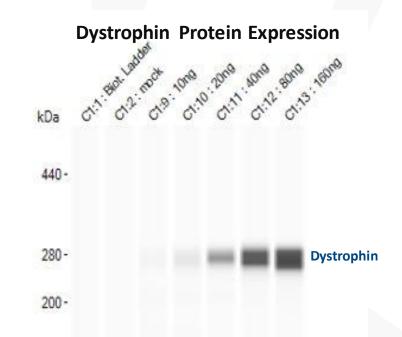


Exon 44 spliced to Exon 56

160ng







Dystrophin protein expressed

Truncated variant



In Vivo POC Study Design

- Objective: Assess muscle function in a humanized, murine model of DMD
- **Test Article:** A single AAV with 2 early generation ARCUS nucleases, expression driven by a muscle specific promoter

• Mouse Models:

- hDMD/mdx *
 - Contains human dystrophin gene and mouse dystrophin gene with mdx mutation
 - Expresses human dystrophin and no mouse dystrophin
- hDMDdel52/mdx **
 - Mutated human and mouse dystrophin gene
 - Does not express functional human or mouse dystrophin

• Readouts:

- Excision of exons 45-55
- Dystrophin restoration
- Force frequency
- Histology for dystrophin⁺ muscle fibers and BaseScope for editing in Pax7⁺ cells

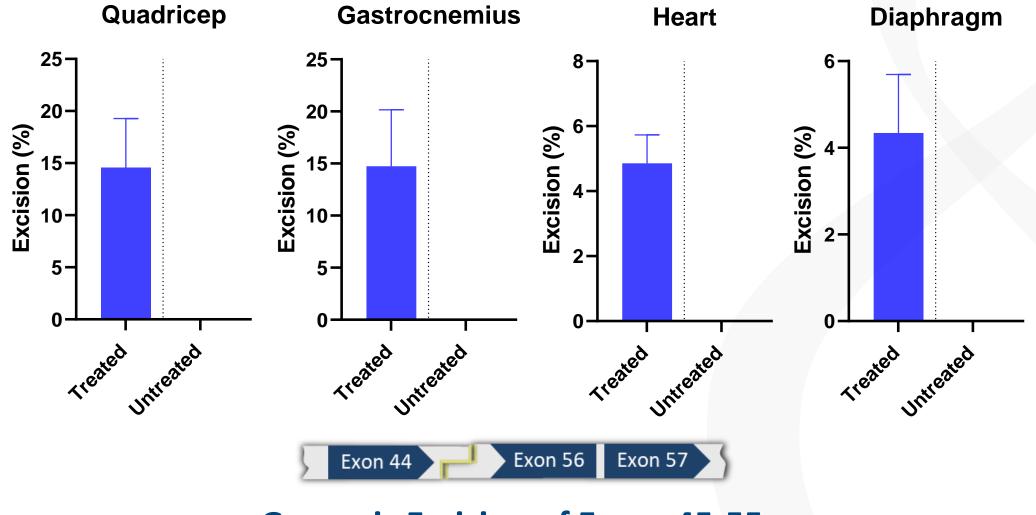
	8 Weeks	
Group	Mouse Strain	Ν
ARCUS-treated	hDMDdel52/mdx	10
Untreated	hDMDdel52/mdx	10
Untreated	hDMD/mdx	10

*'t Hoen PA, de Meijer EJ, Boer JM, Vossen RH, Turk R, Maatman RG, et al. Generation and characterization of transgenic mice with the full-length human DMD gene. J Biol Chem. 2008; 283(9):5899–907. Epub 2007/12/18. https://doi.org/10.1074/jbc.M709410200 PMID: 18083704



**Ya vas A, Weij R, van Putten M, Kourkouta E, Beekman C, Puoliväli J, Bragge T, Ahtoniemi T, Knijnenburg J, Hoogenboom ME, Ariyurek Y, Aartsma-Rus A, van Deutekom J, Datson N. Detailed genetic and functional a nalysis of the hDMDdel52/mdx mouse model. PLoS One. 2020 Dec 23;15(12):e0244215. doi: 10.1371/journal.pone.0244215. PMID: 33362201; PMCID: PMC7757897.

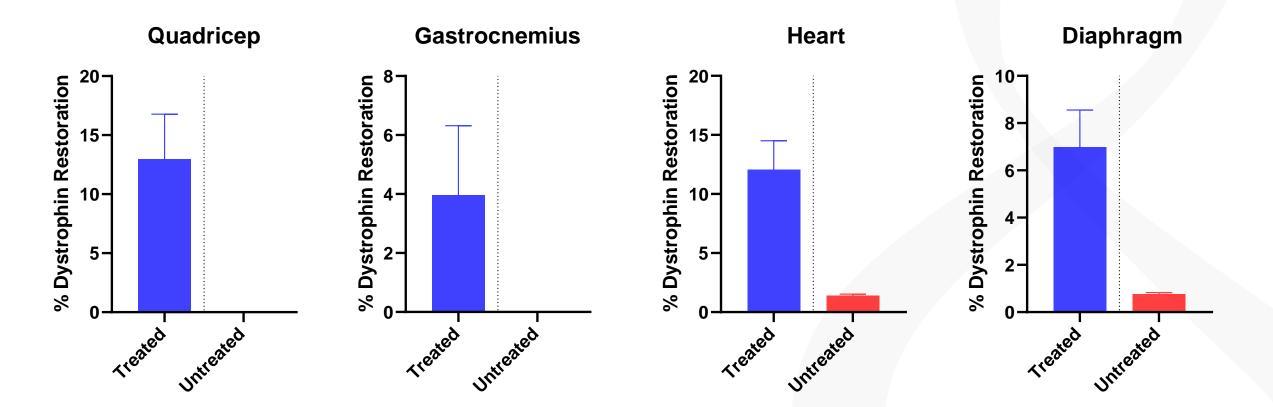
Excision of the "Hot Spot Region" of the Dystrophin Gene in Target Tissues



Genomic Excision of Exons 45-55



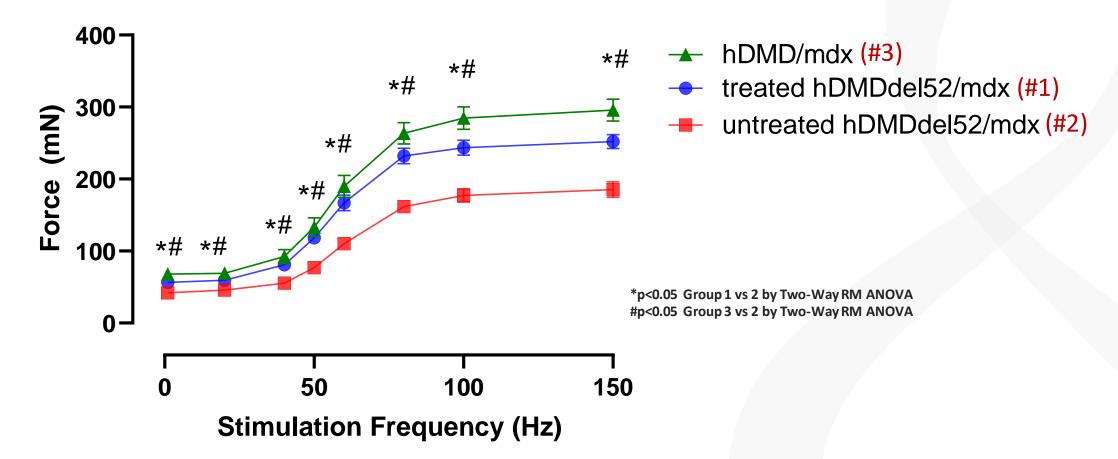
Edited Dystrophin Protein Variant Expressed in Target Tissues



Truncated dystrophin protein produced from splice edited mRNA



Maximum Force Output of Gastrocnemius Muscle in ARCUS-Treated Mice was Significantly Improved

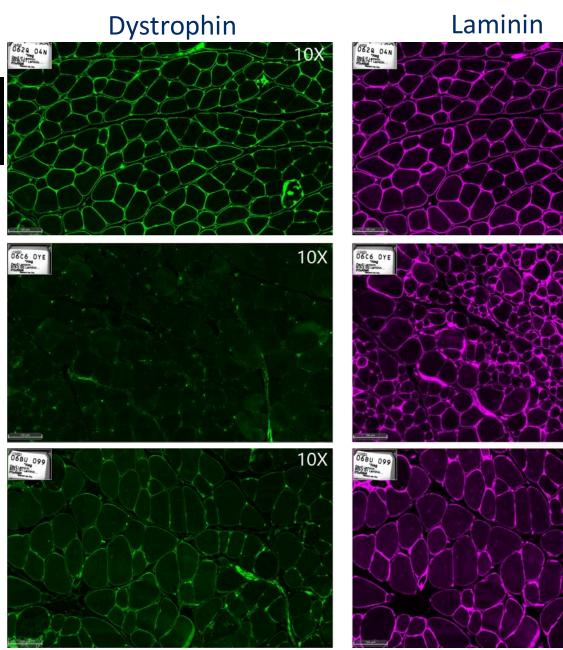


- hDMDdel52/mdx & hDMD/mdx mice were tested 8 weeks post ARCUS AAV delivery
- Changes in muscle function were measured in the gastrocnemius muscle using electrical stimulation and measurement of muscle force
- Maximum force output (MFO) of the gastrocnemius muscle in ARCUS-treated hDMDdel52/mdx mice was significantly improved, reaching 86% of the MFO levels observed in non-diseased, control mice



Restoration of Dystrophin in Gastrocnemius Muscle

Green = Dystrophin Purple = Laminin



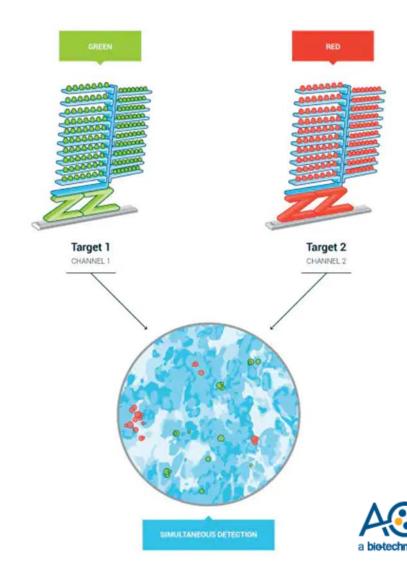
Untreated Non-Disease Model (hDMD/mdx)

Untreated Disease Model (hDMDdel52/mdx)

Treated Disease Model (hDMDdel52/mdx)



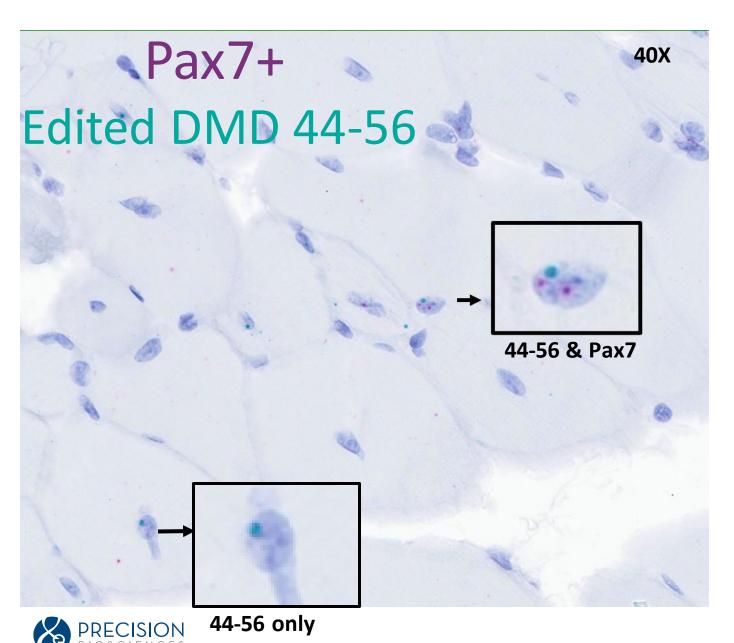
BaseScope[™] Duplex Assay



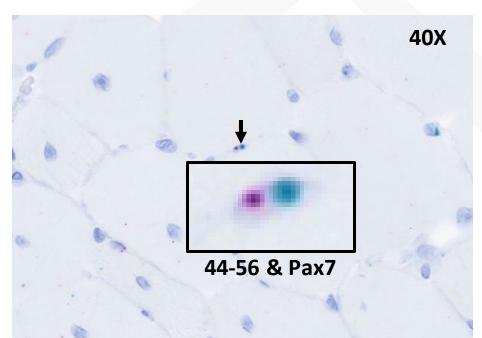
- BaseScope [™] Duplex Assay can be used for simultaneous visualization of two RNA targets while maintaining single cell resolution
- Detection of short RNA target sequences and exon junctions in cells and tissues with morphological context
- Highly specific and sensitive detection of RNA targets with discrimination at the single nucleotide level
- Utilized BaseScope[™] with probes to detect the exon 44-56 splice edited message and Pax7 message, a marker of muscle stem cells (satellite cells)



Edited Dystrophin Transcript in PAX7+ cells, a Marker for Muscle Satellite Cells



- 44-56 Dystrophin mRNA splice edit by BaseScope
- Following AAV delivery, we observed evidence of the edited dystrophin transcript in PAX7+ cells, a marker for muscle satellite cells



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Conclusions

- Here, we report in vivo proof of concept of a dual ARCUS nuclease approach in a humanized DMD mouse model
- Following AAV delivery, we observed the edited dystrophin protein variant in multiple tissue types including heart, diaphragm, and skeletal muscle
- The maximum force output (MFO) of the gastrocnemius muscle in ARCUS-treated animals was significantly improved, reaching 86% of the MFO levels observed in non-diseased, control mice
- Dystrophin protein was restored to muscle fibers with evidence of the edited dystrophin transcript in PAX7⁺ cells, a marker for muscle satellite cells
- This proof-of-concept study demonstrates therapeutic potential of an ARCUS gene editing approach for the treatment of DMD and supports ongoing development toward clinical candidate nomination



Acknowledgements

Precision BioSciences

- Cassie Gorsuch
- Nicole Heard
- Jason Holt
- Derek Jantz
- Matt Jordan-Steele
- Janel Lape
- Whitney Lewis
- Emily Manzon
- Daniel Martin
- Ben Morris
- Dave Morris
- Gary Owens
- Traci Reddick
- Amy Rhoden-Smith
- Kelly Shelton
- Jeff Smith
- Cheng-Wei Wang



Myologica LLC

- Ramzi J. Khairallah
- Jennifer Martin
- Christopher Ward